

Phasic dopamine drives conditioned responding beyond its role in learning

Jay A. Hennig^{1,2,3,4,*}, Mark Burrell^{4,5}, Naoshige Uchida^{4,5}, Samuel J. Gershman^{3,4,6}

¹ Department of Neuroscience, Baylor College of Medicine, Houston, TX, USA

² Neuroengineering Initiative, Rice University, Houston, TX, USA

³ Department of Psychology, Harvard University, Cambridge, MA, USA

⁴ Center for Brain Science, Harvard University, Cambridge, MA, USA

⁵ Department of Molecular and Cellular Biology, Harvard University, Cambridge, MA, USA

⁶ Kempner Institute for the Study of Natural and Artificial Intelligence, Harvard University, Cambridge, MA, USA

* Correspondence: Jay.Hennig@bcm.edu

Competing Interests. The authors have no competing interests to declare.

Acknowledgements. This work was supported by the McNair Foundation (J.H.) and by NIH U19 NS113201 (N.U., S.J.G.).

Abstract

Animals exposed to pairings of a neutral stimulus with reward acquire a conditioned response to the neutral stimulus. A prominent hypothesis, formalized in the Temporal Difference (TD) learning algorithm, is that animals learn to predict the future reward associated with the neutral stimulus (“value”). Though the TD algorithm does not explicitly specify what drives conditioned responding, a typical assumption is that it reflects the animal’s estimate of value. In TD learning, value estimates are updated using reward prediction error (RPE, the discrepancy between observed and predicted reward), and are thought to be signaled by the phasic activity of midbrain dopamine neurons. This hypothesis posits that dopamine’s effects on conditioned responding are mediated entirely by its effects on learning. However, recent experimental and theoretical evidence suggests that dopamine may play a more direct role in modulating conditioned responding. We use a combination of data analysis and computational modeling to probe the relationship between dopamine and conditioned responding. Our results suggest that dopamine directly modulates conditioned responding, in addition to its role in learning. These findings can be captured by a model in which dopamine RPE acts both indirectly (via learning) and directly on conditioned responding.

1 Introduction

2 In Pavlovian conditioning, animals learn to associate a neutral conditioned stimulus (CS) with the delivery
3 of an appetitive or aversive stimulus (US). For example, when a neutral odor or tone cue is repeatedly
4 paired with a subsequent water reward, rodents exhibit anticipatory licking of the water spout during cue
5 presentation. Pavlovian conditioning is often framed through the lens of reinforcement learning theory,
6 which posits that conditioned responding reflects a learned estimate of the cumulative future reward (*value*)
7 following the conditioned stimulus [1, 2, 3]. A leading theory of how animals acquire value estimates is the
8 Temporal Difference (TD) learning algorithm, which updates value estimates using a reward prediction
9 error (RPE), the discrepancy between observed and predicted reward. The RPE is thought to be signaled by
10 the phasic spiking activity of dopamine neurons in the midbrain [4, 5, 6, 7]. This hypothesis is supported by
11 a quantitative match between RPEs and dopamine neuron activity [8, 9, 10, 11], as well as by perturbation
12 experiments establishing the causal role of dopamine in Pavlovian conditioning [12, 13, 14, 15].

13 The dopamine RPE hypothesis holds that dopamine’s role in Pavlovian conditioning is delimited entirely
14 by its role in learning. Any effects dopamine has on conditioned responding must, according to this
15 account, be indirect and delayed. We will challenge this account, presenting data showing that dopamine
16 has a direct, immediate effect on conditioned responding. Several studies already point in this direction
17 [16, 17, 18], though the evidence is mixed [15]. A direct role has been posited by some computational
18 models [19, 20, 21, 22, 23], and might be mediated at the cellular level by the effect of dopamine on the
19 excitability of striatal neurons [24, 25]. However, a systematic empirical investigation of this hypothesis
20 has yet to be undertaken.

21 Disentangling the direct and indirect effects of dopamine on conditioned responding is intrinsically
22 challenging because they cannot be measured separately, and they are often correlated. We address these
23 challenges by conducting a fine-grained analysis of the trial-by-trial covariation between dopamine neuron
24 activity and conditioned responding. Using computational models, we show that the patterns of covaria-
25 tion are most consistent with a model in which dopamine acts both indirectly via learning and directly via
26 modulation of conditioned responding. This model can also account for the heterogeneous effects of optoge-
27 netic perturbations on conditioned responding. Taken together, our findings demonstrate that dopamine’s
28 role in Pavlovian conditioning goes beyond learning, inviting a reconsideration of models positing a direct
29 role in response generation.

30 Results

31 Conditioned responding during contingency degradation covaries with trial-averaged 32 dopamine, and not value

33 We start by revisiting TD learning in the context of trace conditioning (see Methods for mathematical
34 details). In each trace conditioning trial, the animal is presented with a cue (“CS”; e.g., a particular odor),
35 followed by a delay, and then a reward (“US”; e.g., a drop of water; Fig 1A). In a typical TD learning model
36 of trace conditioning, animals learn to estimate the moment-by-moment *value*, or the expected discounted
37 cumulative reward, using RPEs (Fig 1B). The phasic responses of midbrain dopamine neurons to both the
38 CS and US are well described by the RPE signal (see [7] for a review), supporting their hypothesized role

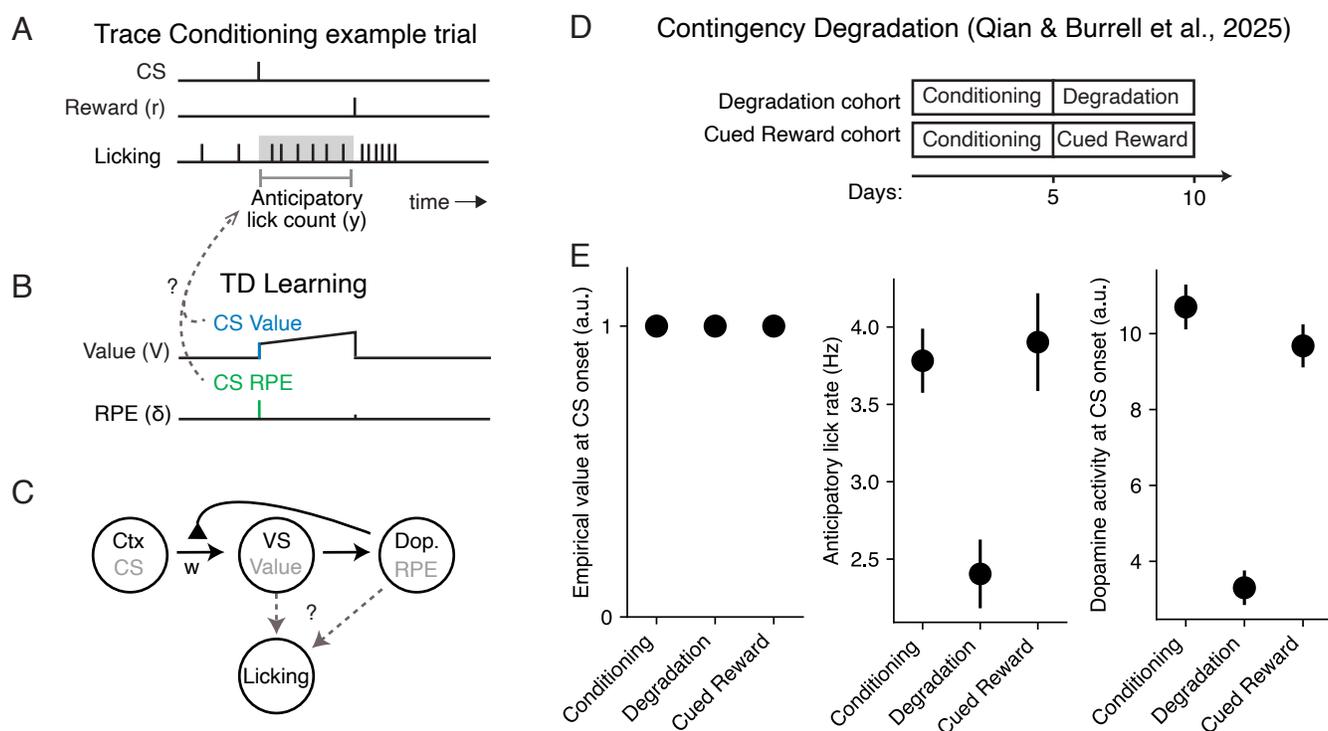


Fig 1. TD learning and conditioned responding during trace conditioning with contingency degradation. **A.** Example trace conditioning trial consisting of a conditioned stimulus (CS), followed by a delay, and then a reward stimulus. Also depicted is an example of the times the animal licked the reward spout, with anticipatory licking (the conditioned response) measured during the interval between the CS onset and reward delivery. **B.** Example of the estimated value and reward prediction error (RPE) of TD learning during the example trial shown in panel A. Dashed arrows depict different potential relationships between CS Value, CS RPE, and anticipatory licking. **C.** Putative circuit implementation of TD learning, where sensory cortex provides the CS input, ventral striatum (VS) carries the value estimate, and midbrain dopamine neurons provide the RPE signal, which modulates feedforward plasticity between cortex and VS. **D.** Experimental structure of the contingency degradation experiment [26]. Two cohorts underwent a “Conditioning” phase for five days/sessions, followed by five days of either a “Degradation” or “Cued Reward” phase for the Degradation and Cued Reward cohorts, respectively. **E.** Average objective value at CS onset, anticipatory lick rate, and dopamine activity (z-scored axonal calcium fluorescence) at CS onset, for the Odor A cue during each of three phases shown in panel D. Black circles and lines denote mean \pm SE across trials.

39 in TD learning and the interpretation of dopamine activity as a putative measure of the animal’s RPE.
 40 Mice trained on trace conditioning with an odor CS and a water reward produce anticipatory lick-
 41 ing during the delay period—their conditioned response (Fig 1A). To link the TD model to conditioned
 42 responding, we require an auxiliary assumption about response generation (Fig 1C). Conditioned respond-
 43 ing typically covaries with the CS value. However, as we’ll see, differences in CS RPEs across cues and
 44 across time are also correlated with conditioned response rates. This raises the possibility that conditioned
 45 responding could additionally reflect CS RPEs, putatively signaled by phasic dopamine.
 46 Recent work using a contingency degradation paradigm supports this possibility [26]. In this study,
 47 there were three distinct experimental phases, presented to two different cohorts of mice (Fig 1D). Briefly,
 48 during the Conditioning phase, each trial consisted of a CS (“Odor A”) followed by a variable reward. Mice
 49 in the Degradation cohort experienced a Degradation phase, where Odor A trials were interleaved with
 50 trials containing uncued rewards. Mice in the Cued Reward cohort experienced a Cued Reward phase,
 51 where Odor A trials were interleaved with reward trials cued by an additional odor CS. Critically, the Odor

52 A CS was rewarded similarly across all three phases, meaning its objective value was held constant across
53 every phase (Fig 1E, left). Notably, animals' anticipatory licking was decreased in the contingency phase
54 compared to the conditioning and cued reward phases (Fig 1E, middle), even though the CS value was
55 constant across phases. This mirrored the CS dopamine response (Fig 1E, right), and was well explained by
56 a TD learning model where CS dopamine reflects the CS RPE. This suggests that differences in conditioned
57 responding may be driven by CS RPEs and not CS value.

58 While the possibility that conditioned responding is based on CS RPE and not CS value is suggestive,
59 the evidence presented in [26] is based on trial-averaged activity and correlations across cues or phases of
60 trials. If the CS RPE is truly driving conditioned responding, then the correspondence between dopamine
61 and licking should also manifest on a trial-to-trial basis for a single cue.

62 **Trials with larger CS dopamine have higher conditioned response rates**

63 To look for evidence of a trial-to-trial correspondence between CS dopamine and conditioned responding,
64 we first compared the number of anticipatory licks with the peak magnitude of dopamine activity following
65 CS onset. We first carry out this analysis on each trial of an example session at the end of conditioning for
66 one mouse, and then show the results aggregated across all sessions and mice. For this and all following
67 analyses of data from [26], we only consider trials with Odor A, which is the CS that was identically
68 rewarded throughout the experiment (i.e., the objective value of this CS was constant). "Dopamine
69 activity" here means the z-scored axonal calcium fluorescence (see Methods). We labeled trials where
70 the peak CS dopamine activity was in the highest quartile across trials as a "High CS dopamine" trial (Fig
71 2A), and those where the peak in CS dopamine activity was in the lowest quartile as a "Low CS dopamine"
72 trial (Fig 2B). In this example session (Fig 2C), trials with high CS dopamine had more anticipatory licks
73 on average than trials with low CS dopamine (Fig 2C; one-sided Wilcoxon rank sum test, $W = 2.638$,
74 $p = 0.004$).

75 This same general pattern was also evident across sessions from all mice, where we observed that trials
76 with high CS dopamine had higher anticipatory lick rates than did trials with low CS dopamine (Fig 2D;
77 one-sided Wilcoxon rank sum test, $W = 7.561$, $p < 1 \times 10^{-3}$). This effect could not be explained by
78 differences in satiety, as high CS dopamine trials had higher anticipatory lick rates than low CS dopamine
79 trials even when excluding the last half of trials from each session (one-sided Wilcoxon rank sum test,
80 $W = 3.769$, $p < 1 \times 10^{-3}$). CS dopamine and anticipatory lick rates had a graded relationship, such that
81 higher CS dopamine was correlated with larger anticipatory lick rates on the last day of conditioning for
82 every mouse (Fig 2D; average Pearson correlation was 0.33 ± 0.05 , mean \pm SE; $N = 14$ animals). While TD
83 learning predicts that both CS dopamine and CS value should increase with learning, this correspondence
84 between anticipatory licking and CS dopamine is unexpected on the last day of conditioning, when animals
85 have fully learned the reward contingencies. This result is also consistent with empirical results in previous
86 studies that found anticipatory lick rates were decreased when dopamine neurons were transiently inhibited
87 during the ISI using optogenetics [18, 15], a result we will return to in a later section.

88 Differences in the average lick rates of rodents may be due to differences in licking latency, intensity, or
89 duration [27, 28, 29, 30, 31]. We found that trials with high CS dopamine did have slightly shorter latencies
90 to the time of the first lick than trials with low CS dopamine (Fig 2F; one-sided Wilcoxon rank-sum test,
91 $W = -4.374$, $p < 1 \times 10^{-3}$). Anticipatory lick rates were also larger on high versus low CS dopamine

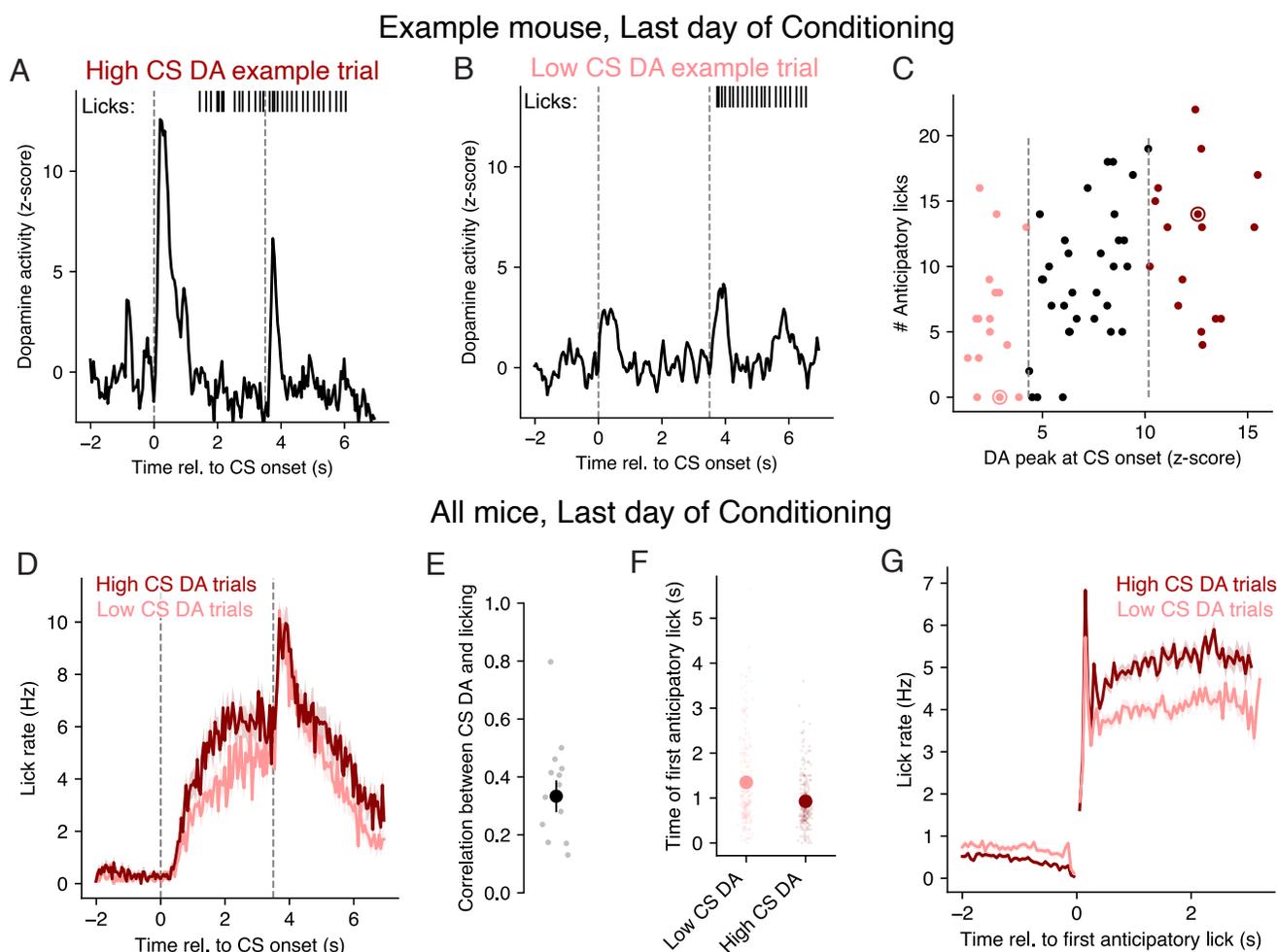


Fig 2. Anticipatory lick rates late in conditioning are larger on trials with larger CS-evoked dopamine. **A-B.** Dopamine activity (black trace) and licks (black vertical lines) during an example “High CS dopamine” (panel A) and “Low CS dopamine” (panel B) trial, on the last day of conditioning from an example mouse. **C.** Number of anticipatory licks and dopamine response to the CS on trials from the same example session as panels A and B. Dashed lines depict lower and upper quartiles of the CS dopamine response, used to define “Low CS dopamine” (light red) and “High CS dopamine” (dark red) trials, respectively. Dark red and light red circles indicate the example trials shown in panels A and B, respectively. **D.** Average lick rate (mean \pm SE) across all High CS dopamine and Low CS dopamine trials from the last day of conditioning in all mice. **E.** Pearson correlation between anticipatory lick rate and CS dopamine across trials in the last day of conditioning for each mouse (gray dots), and across mice (black circle). Black line depicts mean \pm SE. **F.** Average time of the first anticipatory lick (mean \pm SE) across the same trials as in panel D. **G.** Average lick rate (mean \pm SE) after aligning lick times within each trial to the time of the first anticipatory lick.

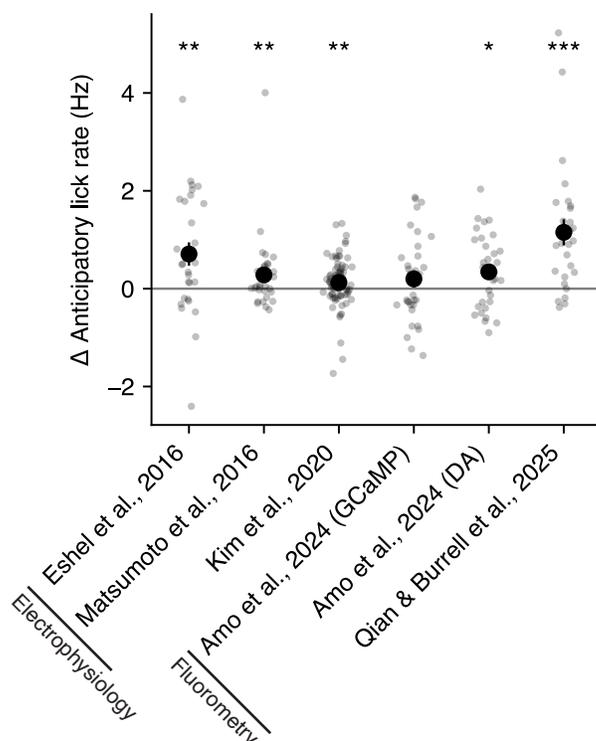


Fig 3. Anticipatory lick rates are larger on trials with higher CS dopamine than on trials with low CS dopamine across multiple studies. For each study, gray dots depict the average difference in the anticipatory lick rate on trials with High versus Low CS dopamine within a single session. Black circles depict the mean across sessions, and black lines depict the mean \pm SE across sessions. Asterisks depict significance of a Wilcoxon signed rank test at the 0.05 (*), 0.01 (**), and 0.001 (***) level. Black lines on x-axis tick labels indicate the dopamine signal recording technique used in that study. For [33], results are shown separately for dopamine activity measured using calcium (GCaMP) and dopamine (DA) sensors.

92 trials when aligning to the time of the first lick (Fig 2G; one-sided Wilcoxon rank-sum test, $W = 12.902$,
93 $p < 1 \times 10^{-3}$). Thus, trials with high CS dopamine had both shorter licking latencies and higher lick rates
94 than trials with low CS dopamine.

95 We next asked whether these differences in lick rate as a function of the CS dopamine response were
96 present in other studies that recorded both dopamine and licking in mice undergoing odor trace conditioning
97 [10, 32, 11, 33]. While the effect size varied across studies, trials with high CS dopamine all tended to
98 have higher rates of anticipatory licking than trials with low CS dopamine across a range of studies and
99 dopamine signals (Fig 3). These results suggest that the magnitude of the CS dopamine response can, on
100 average, explain variability in the subsequent amount of anticipatory licking. As we will show in a later
101 section, this finding cannot be explained by the traditional TD learning assumption that CS value solely
102 drives conditioned responding.

103 CS dopamine and conditioned responding covary throughout conditioning

104 We next asked whether the relationship between CS dopamine and anticipatory licking observed at the
105 end of conditioning was also present throughout the learning process. We visualized the time series of CS
106 dopamine and anticipatory licking on each trial to the same cue during the conditioning phase of [26]. CS
107 dopamine and anticipatory lick rates were positively correlated across all animals (Fig 4), with an average

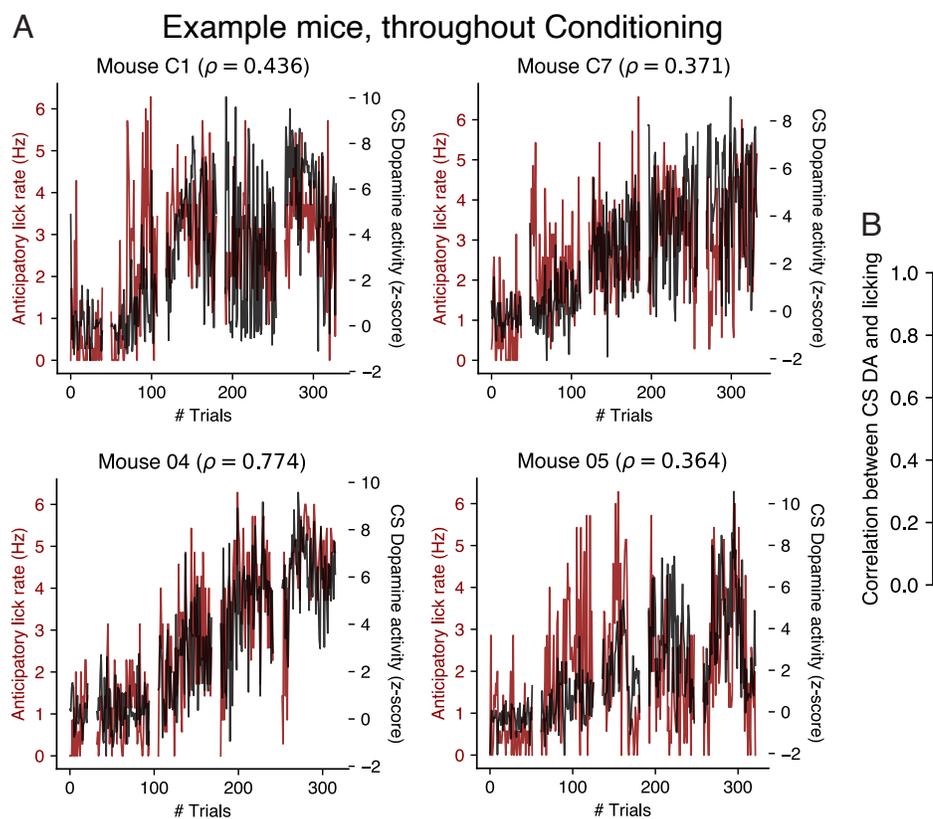


Fig 4. Trial-by-trial correlations between anticipatory lick rates and CS-evoked dopamine, throughout conditioning. **A.** Example time courses of anticipatory lick rate (red) and CS dopamine response (black) to the same cue on each trial of conditioning, with gaps of 10 trials added between sessions for visualization of session boundaries. Pearson correlation (ρ) between anticipatory lick rate and CS dopamine is shown above each subpanel. **B.** Pearson correlation between anticipatory lick rate and CS dopamine across conditioning trials for each mouse (grey dots), and across all mice (black circle). Black line depicts mean \pm SE.

108 Pearson correlation of 0.44 ± 0.04 (mean \pm SE; $N = 14$ animals). While a TD learning agent’s estimate of CS
109 value and its CS reward prediction error are expected to monotonically increase with learning, empirical
110 measures of anticipatory licking and CS dopamine (assumed to reflect the CS reward prediction error)
111 showed substantial non-monotonicity, suggesting this positive correlation may not be solely due to learning.
112 Overall, our results suggest that trial-to-trial variability in CS dopamine can partially explain trial-to-trial
113 variability in conditioned responding. As we will show next, this correspondence is *not* expected from a
114 TD learning agent whose rates of conditioned responding reflect CS value.

115 **Anticipatory licking is maximally correlated with the immediately prior CS dopamine** 116 **response**

117 One caveat in interpreting the correlations between CS dopamine and licking above is that, even if licking
118 were driven solely by value, TD learning would still predict correlations between the CS RPE (i.e., CS
119 dopamine) and licking. This is because standard TD learning models predict that the RPE will gradually
120 backpropagate across trials from the reward to the CS [4], as observed empirically [6]. This has been
121 described as an ‘indirect’ relationship between the RPE/dopamine and responding [19], because the CS
122 value on each trial will in general correlate with the RPEs from *previous* trials. Alternatively, the CS
123 RPE may have a ‘direct’ (i.e., simultaneous) role in responding, in which case we would expect that
124 the anticipatory response on trial t will depend maximally on the CS RPE on trial t . Thus, positive
125 correlations between the CS RPE and anticipatory licking could, in principle, be explained both by the
126 traditional assumption that licking reflects CS value, or by a model where licking is modulated directly by
127 CS RPE.

128 To understand whether both of these hypotheses are indeed consistent with empirical data, we took
129 a “phenotyping” approach (Fig 5A). First, we simulated thousands of TD learning agents, where each
130 model had randomly sampled hyperparameters (see Methods). Next, we generated time series of anticipa-
131 tory licking according to two different hypotheses. Under the *indirect* dopamine hypothesis (H1), licking
132 is a readout of CS value. Under the *direct* dopamine hypothesis (H2), licking is a readout of the CS
133 RPE/dopamine. For each resulting agent, we then assessed the Pearson correlation between the number
134 of anticipatory licks on trial t and the magnitude of the CS RPE on trials $t - \tau$ for $\tau = 0, 1, \dots, 4$. We
135 refer to the set of resulting correlations, $\beta \in \mathbb{R}^5$, as the agent’s *phenotype*.

136 We found that the resulting phenotypes differed based on which agents had licking generated by CS
137 value or by CS RPEs (Fig 5B). Agents whose licking directly reflected the CS RPE were distinguished by
138 a positive correlation between anticipatory licking and the CS RPE on the same trial (i.e., $\tau = 0$) (Fig
139 5B, green traces). Overall, agents whose licks reflected H2 versus H1 had larger correlations for $\tau = 0$
140 ($d' = 1.751 \pm 0.077$, mean \pm s.d. across $N = 1,000$ bootstraps). Thus, this phenotyping approach could
141 distinguish between two classes of TD agents where licking was driven either directly or indirectly by the
142 RPE.

143 We next applied this same phenotyping approach to empirical data from all of the trace conditioning
144 studies we considered earlier (see Methods). We found that anticipatory licking on a given trial was
145 significantly positively correlated with the CS dopamine on the same trial across all previous studies (Fig
146 5C). This supports the idea that CS RPE/dopamine plays a direct role in driving conditioned responding
147 during this experiment.

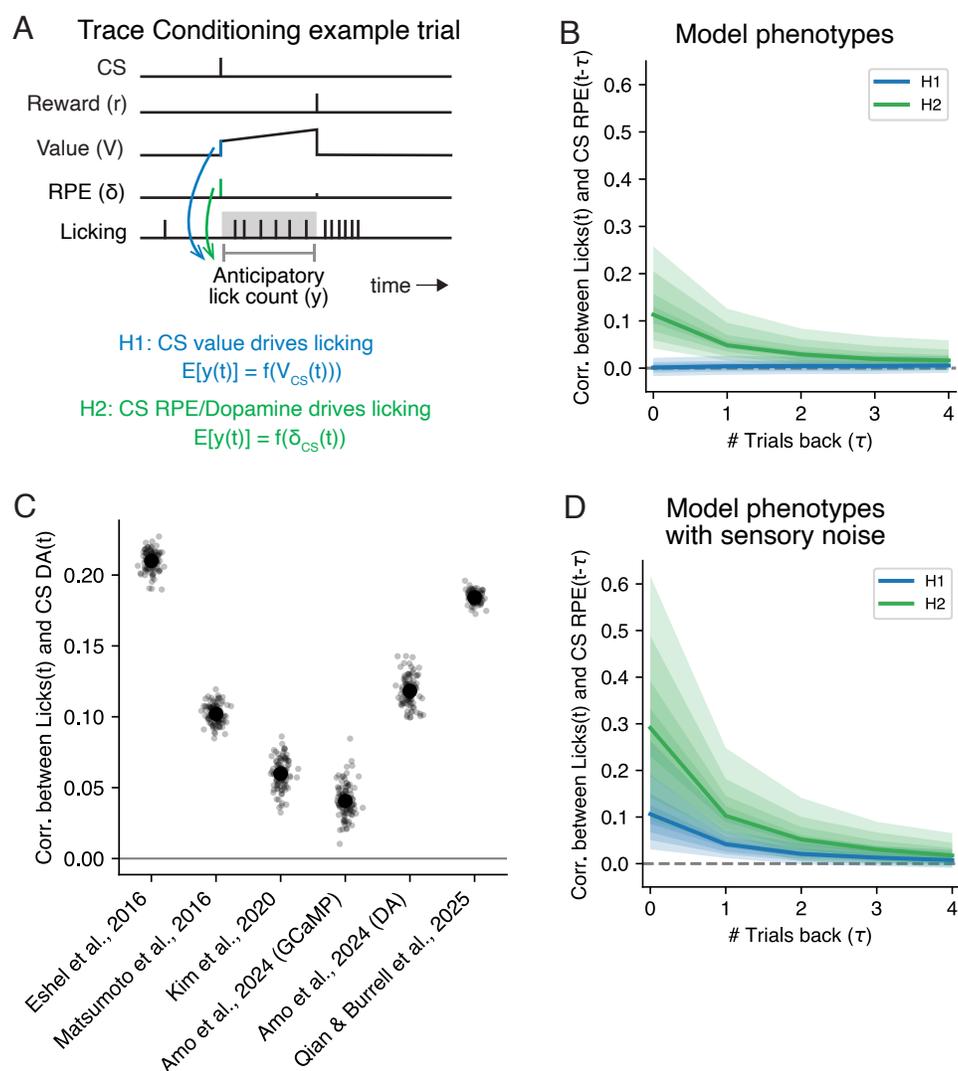


Fig 5. Phenotyping suggests anticipatory licking depends directly on CS RPEs/dopamine, or CS value with sensory noise. **A.** Schematic depicting two hypotheses for how the anticipatory lick count on a given trial, $y(t)$, might depend on either CS value ($V_{CS}(t)$; H1) or on the CS RPE ($\delta_{CS}(t)$; H2). **B.** Correlation between anticipatory lick count and CS RPE on previous trials, for simulated TD agents whose licking was generated according to H1 (blue) or H2 (green). Lines depict mean correlations across $N = 500$ models, and shading depicts deciles across models. **C.** Pearson correlations between anticipatory lick count and CS dopamine on the same trial, for empirical measurements from previous studies. Black dots indicate correlations across bootstraps, while black circles and lines indicate mean \pm SE across $N = 1,000$ bootstraps. All correlations were statistically significant compared to correlations calculated using trial-shuffled DA. **D.** Same as panel B, but for agents with added sensory noise.

148 A positive correlation between CS dopamine and licking does not imply that the two are causally
 149 related. We reasoned that positive correlations could also arise if licking reflects CS value, but both CS
 150 value and licking have shared input noise. To implement this idea, we added noise to the CS sensory
 151 representation (e.g., reflecting a noisy sensory response to the odor) on each trial before applying TD
 152 learning, and generated licking according to the indirect model where licking reflects CS value. This
 153 addition of sensory noise produced similar, though weaker correlations between the CS RPE and licking
 154 on the same trial (Fig 5D). Nevertheless, the first regression weight was still consistently larger for agents
 155 whose licks reflected H2 versus H1 ($d' = 1.245 \pm 0.060$, mean \pm s.d. across $N = 1,000$ bootstraps).

156 Overall, our analyses revealed that, across a range of conditioning experiments, licking and CS dopamine
157 were strongly positively correlated on the same trial, as expected by a TD model where CS RPEs (putatively
158 signaled by dopamine) directly drive licking. Our phenotyping approach suggests that this correlation
159 between CS dopamine and anticipatory licking on the same trial may be due either to a direct role for CS
160 dopamine in licking, or to the presence of substantial noise in the CS sensory representation. Empirical
161 support of the former possibility will be evaluated in a later section.

162 **Uncued peaks in CS dopamine between trials precede changes in licking**

163 The analyses above revealed a correspondence between the concurrent CS RPE and conditioned responding.
164 Our modeling suggests that this could be explained either by dopamine having a direct role in modulating
165 licking, *or* a value-licking model with substantial noise in the sensory representation of the cue. One reason
166 that these two possibilities are hard to distinguish is that both the CS RPE and CS value are directly
167 dependent on the CS sensory representation. We reasoned that, if dopamine can directly modulate licking,
168 then peaks in dopamine that are *not* cue-evoked may also yield changes in licking.

169 We therefore turned to comparing dopamine and licking during the intertrial interval (ITI) of data from
170 [26], an epoch containing no experimentally-controlled inputs (Fig 6). Dopamine activity often showed
171 substantial phasic activity during the ITI (Fig 6A), and lick rates were significantly larger in the 250 msec
172 after versus before each peak (Fig 6B; Wilcoxon signed rank test $T = 86.0$, $p < 1 \times 10^{-3}$). We defined a
173 “peak” in dopamine activity during the ITI as a time during the ITI at which the dopamine signal surpassed
174 more than 3 standard deviations above average background levels across the entire experiment. We then
175 aligned licking to each of these peaks and averaged across each peak, resulting in a “dopamine-peak-
176 triggered average” of licking during the ITI. This analysis revealed an abrupt increase in licking following
177 a dopamine peak during the conditioning phase (Fig 6C). This increase in licking was dose-dependent: the
178 magnitude of these uncued dopamine peaks was positively correlated with larger subsequent increases in
179 licking (Fig 6D). Importantly, because these dopamine peaks occurred during the ITI, they were uncued,
180 and thus had zero objective value (because they were not predictive of reward). Nevertheless, these
181 dopamine peaks preceded increases in licking, suggestive of a direct role for dopamine in modulating
182 anticipatory licking.

183 To quantify this result for each session from each mouse, we compared the number of licks immediately
184 before versus after the time of each dopamine peak, and summarized this difference using the sensitivity
185 index (d' ; see Methods). We then visualized the d' of each cohort throughout the conditioning, degradation,
186 and cued reward phases (Fig 6E-F). Across sessions of both cohorts, d' was significantly positive throughout
187 conditioning (Fig 6E-F; two-sided Wilcoxon signed rank test $T = 160.0$, $p < 1 \times 10^{-3}$), meaning the number
188 of licks was consistently larger directly after versus before an uncued peak in dopamine during the ITI.
189 For mice in the Contingency Degradation phase (i.e., some trials contained uncued rewards), d' was also
190 significantly positive (Fig 6E; two-sided Wilcoxon signed rank test $T = 7.0$, $p < 1 \times 10^{-3}$). However, for
191 mice in the Cued Reward phase, d' was no longer positive (Fig 6F; two-sided Wilcoxon signed rank test
192 $T = 98.0$, $p = 0.083$). This result, where uncued peaks in dopamine preceded increases in licking during
193 the ITI for some experimental conditions but not others, suggests a nonstationary or dynamic relationship
194 between uncued peaks in dopamine and licking. While dopamine may directly modulate anticipatory
195 licking, this relationship may be gated by other signals related to reward expectation, a possibility we will

206 We first considered results from a study that used optogenetics to transiently excite dopamine neurons
207 during cue onset [17] (Fig 7A). The experimenters found that if dopamine neurons were stimulated during
208 CS delivery, anticipatory licking was high for cues paired with reward (“CS+ Excitation”), but remained
209 low for cues *not* paired with reward (“CS- Excitation”). This suggests that dopamine stimulation on its
210 own was not sufficient to cause licking.

211 We applied TD learning models to these experiments, modeling dopamine excitation as a constant, b ,
212 added to the RPE, δ , simultaneous with the cue delivery (see Methods). We then simulated anticipatory
213 licking as a readout of either CS value (Fig 7B) or CS RPE (Fig 7C). These are the same two models we
214 considered earlier in Fig 5. While the results in [17] could be reproduced by TD learning models where CS
215 value directly drove licking (Fig 7B), these results were also explained by a model where CS RPEs directly
216 drove licking (Fig 7C). Notably, though the latter model has a direct link between the CS RPE and licking,
217 pairing dopamine stimulation with cue presentation did *not* lead to large lick rates for the unrewarded
218 CS- cue. This is because the CS RPE itself depends on CS value, which remains low for unrewarded cues.
219 Thus, even TD models with a direct link between the CS RPE/dopamine and licking can nevertheless
220 show a “gated” response to dopamine excitation. Because this result from [17] was explainable with both
221 models of licking, these empirical results do not speak to whether or not the CS RPE has a direct role in
222 anticipatory licking.

223 We next considered results from a pair of studies that used optogenetics to phasically *inhibit* dopamine
224 activity during the cue or reward onset [15, 18] (Fig 7D). The authors found that dopamine inhibition
225 during either the cue or the reward onset reduced anticipatory lick rates, with inhibition during the reward
226 onset having a larger relative impact. We modeled dopamine inhibition as a constant, b , subtracted from
227 the RPE/dopamine signal, δ , simultaneous with either the cue or reward delivery. We then applied TD
228 learning models to these experiments as before (see Methods). We simulated anticipatory licking on each
229 trial using either the CS value (Fig 7E) or CS RPE (Fig 7F) on each trial. Again we found that both models
230 could reproduce the empirical results [15, 18], where lick rates were reduced for dopamine inhibition during
231 either the cue or the reward onset, but with reward inhibition having a larger impact. Thus, these results
232 do not speak to whether or not the CS RPE directly modulates anticipatory licking.

233 One reason why exciting or inhibiting dopamine can impact anticipatory licking even if CS RPE does
234 not directly drive licking is due to the fact that these perturbations were applied in blocks of adjacent trials.
235 These blockwise perturbations allow perturbed dopamine levels to impact CS value, and therefore licking,
236 through learning. A more direct way of evaluating whether the CS RPE has an immediate influence on
237 licking would be excite or inhibit dopamine activity on *random* trials rather than in blocks. Indeed, for TD
238 models where licking was determined solely by CS value, randomly inhibiting CS RPE on random trials did
239 *not* lead to reduced anticipatory licking compared to trials where CS dopamine was not inhibited (Fig 7G,
240 top). By contrast, anticipatory licking was reduced during inhibited trials for TD models where licking was
241 determined by CS RPE (Fig 7G, bottom). In fact, previous work did find that, when inhibiting dopamine
242 during the cue period of random 50% of trials, anticipatory lick rates were lower on the randomly inhibited
243 dopamine trials than on the uninhibited dopamine trials [15]. Thus, this empirical result combined with
244 our simulations supports the idea that CS RPEs/dopamine can directly modulate anticipatory licking on
245 the same trial, and that this modulation is distinct from the gradual changes in licking expected due to
246 learning.

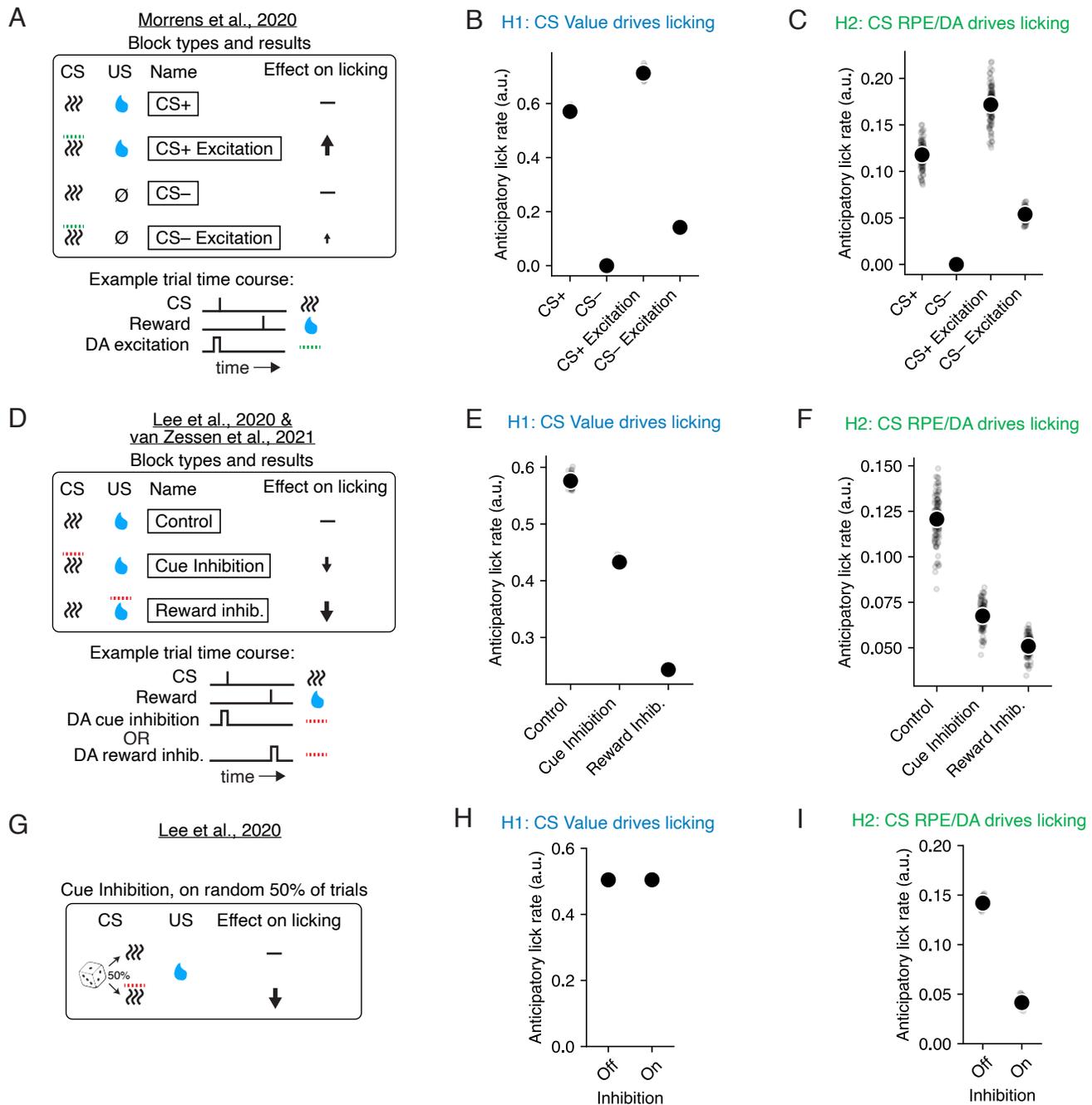


Fig 7. A model where the CS RPE drives anticipatory licking can explain the effects of causal manipulations of dopamine on licking. **A.** Top, four distinct conditions from [17], each with a distinct CS. “CS+” indicates a deterministically rewarded cue, while “CS-” indicates a deterministically unrewarded cue. For CS+ and CS- Excitation trials, optogenetic excitation of dopamine was modeled as a phasic, additive increase in the RPE during the CS delivery (see Methods). Anticipatory licking was high for CS+ Excitation and low for CS- Excitation. Bottom, example trial time course of a CS+ Excitation trial. **B.** Average lick rate for each trial type (each simulated separately), for models where lick rate is proportional to CS value (see Methods), across 100 experiments (black circles and lines indicate mean \pm SE for $N=100$ experiments). **C.** Same as panel B, but where lick rate is proportional to CS dopamine (see Methods). **D.** Top, three conditions and results from [15] and [18], each with a distinct CS. Optogenetic inhibition of dopamine was modeled as the opposite of excitation. Anticipatory licking was lowest in the Reward inhibition condition, but still reduced for Cue inhibition. Bottom, example trial time course of Cue inhibition versus Reward inhibition trials. **E-F.** Same conventions as panels B-C, but for the conditions in panel D. **G.** In a separate condition of [15], dopamine cue inhibition was either applied (“On”) or not applied (“Off”) during the delivery of a single CS on a random 50% of trials. Lick rates were reduced on the trials with cue inhibition. **H-I.** Same conventions as panels B-C, but for the conditions in panel G.

247 Discussion

248 Using a combination of data analysis and computational modeling, we probed the relationship between
249 phasic dopamine and anticipatory licking in mice during Pavlovian conditioning. Using data from multiple
250 previously published studies, we found that the response of dopamine neurons to a reward-predictive cue
251 (“CS dopamine”) was positively correlated with a higher anticipatory lick rate on the same trial. This
252 suggests a link between the CS dopamine and the immediate rate of conditioned responding, a relationship
253 not expected from the standard assumption that responding is proportional to CS value. In addition,
254 we found that positing a direct link between the CS dopamine and anticipatory licking was necessary to
255 reproduce previous empirical results showing that optogenetically inhibiting CS dopamine on random trials
256 led to a decrease in anticipatory licking on those same trials. Taken as a whole, our results suggest that
257 phasic dopamine directly modulates conditioned responding in addition to its canonical role as a learning
258 signal. This direct role can be captured by a model in which the CS RPE drives conditioned responding.

259 The possibility that dopamine plays a role in action selection, and therefore conditioned responding, has
260 been considered by previous computational models, mostly from the perspective of “incentive salience”—
261 i.e., the role of dopamine in assigning motivational value to objects or actions [19, 22, 20, 35]. Previous
262 studies using Pavlovian conditioning in rats have found that some rats develop “sign-tracking” behavior,
263 where reward-predictive cues develop incentive motivational value, or incentive salience. This sign-tracking
264 behavior is known to be dependent on dopamine [36, 20, 37, 38]. Incentive salience in tasks such as the
265 ones we consider here, using odor cues, has not been studied to our knowledge. Previous work has found
266 that dopamine release in the ventral striatum can initiate sniffing via action on both D1- and D2-expressing
267 medium spiny neurons (MSNs) [39]. Sniffing could be one way of measuring sign-tracking of odor cues, in
268 which case it would be interesting to evaluate the connection between trial-by-trial differences in sniffing,
269 licking, and CS dopamine.

270 One potential circuit implementation consistent with a direct role for the CS RPE/dopamine in re-
271 sponding is if phasic dopamine provides feedforward excitation of striatal medium spiny neurons (MSNs).
272 There is some empirical support of this idea [25], though this remains debated [40]. D1-MSNs are thought
273 to modulate lick rate [41]. The response of MSNs to reward-predictive cues, putatively encoding CS
274 value, may be modulated in a feedforward manner by CS dopamine, putatively encoding the CS RPE.
275 Computational models based on this idea have been considered previously [42, 21].

276 Our findings add to a growing body of work on the role of phasic dopamine in movement vigor and ini-
277 tiation in other paradigms, spanning operant conditioning [42, 43, 44, 45], Pavlovian-instrumental transfer
278 [46], and cue-potentiated feeding [27]. Closely related to our findings is work identifying a role for fast
279 timescale changes in phasic dopamine activity in self-timed or self-initiated movements [47, 48, 49]. Our
280 approach complements these studies by considering the relationship between dopamine and responding in
281 the context of dopamine as the RPE signal in TD learning, which allows us to dissect the contributions
282 of dopamine on responding that are immediate versus gradual through learning. An important direction
283 for future work is to develop a unified computational framework that can explain the role of dopamine on
284 both the timing and vigor of movements across these different paradigms.

285 Previous studies have considered the possibility that CS dopamine plays a direct role in modulating
286 responding during Pavlovian conditioning. In [17], the authors dismissed this possibility because “CS
287 dopamine stimulation in our experiments did not cause licking, per se, but selectively promoted responding

288 only to rewarded cues.” As we showed in Fig 7A-C, however, this result on its own is not sufficient to
289 discriminate between a TD-based model where licking is directly modulated by CS value or CS RPE. Fully
290 discriminating between the roles of CS value and CS RPEs in modulating responding is challenging because,
291 while we can measure putative RPEs using dopamine activity as a proxy, we cannot directly measure CS
292 value. Another challenge is that causal perturbations to dopamine presented in the literature are typically
293 applied in blocks rather than on randomly selected trials. This makes it challenging to distinguish between
294 the learning and feedback-related effects of dopamine expected over the course of multiple trials, versus the
295 immediate/feedforward effects expected on a single trial. The only study we are aware of that randomly
296 perturbed dopamine activity during a Pavlovian trace conditioning task is [15], which we have shown here
297 provides strong evidence that CS dopamine can directly modulate responding.

298 The slow correlations we observed between CS dopamine and conditioned responding in Fig 4 suggests
299 that variability in an animal’s motivation or arousal may underlie some of the co-fluctuations in dopamine
300 and licking. While arousal cannot explain the impacts of random optogenetic dopamine inhibition on licking
301 (Fig 7G), arousal may nevertheless drive some co-fluctuations in both dopamine and licking. Previous work
302 suggests a correspondence between conditioned responding and pupil size (an index for arousal) during
303 trace conditioning tasks [50], but how and whether animals’ arousal differentially modulates CS dopamine
304 and responding is an interesting, unanswered question.

305 The relationship between RPE and conditioned responding brings our work into contact with a broader
306 set of ideas in animal learning theory [51, 52, 53, 54]. These accounts draw an important distinction between
307 the number of training trials until a response acquisition criterion is met (sometimes called *associability*)
308 and terminal response rate (after extensive training). It has been noted that reward expectation on its
309 own is an inadequate predictor of associability. Rather, it is the *difference* between ISI (cue) and ITI
310 (contextual) reward expectations that matters. The example of contingency degradation, discussed above,
311 illustrates this point well: animals will not learn to produce conditioned responses to reward-predictive
312 cues if the rate of reward is the same in the ISI and ITI [55]. This contrastive view of associability fits
313 naturally with the contrastive nature of RPEs. However, here we have focused on the role of dopamine in
314 driving terminal response rates, not associability, which we leave to future work.

315 In conclusion, our findings suggest that RPEs, putatively signaled by phasic dopamine activity, may
316 directly modulate the intensity of conditioned responding on a trial-by-trial basis. This conclusion is
317 supported by converging correlational evidence across multiple datasets, as well as the effects of random
318 optogenetic stimulation of dopamine during cue presentation. Future work disentangling the contributions
319 of sensory noise and arousal, as well as how dopamine could drive responding at the circuit level, will be
320 important for building a more complete picture.

321 **Methods**

322 **Analysis of dopamine activity and anticipatory licking**

323 We analyzed previously collected dopamine and licking activity recorded from multiple trace conditioning
324 studies conducted with mice [10, 32, 11, 33, 26]. For all studies, dopamine activity was analyzed in 50 msec
325 bins. CS dopamine activity was defined using the activity within a 500 msec window following CS onset.
326 For studies that recorded dopamine using electrophysiology [10, 32, 11], we used the average firing rate

327 within the window. For studies that used fluorometry [33, 26], we used the maximum dopamine activity
328 within the window. The anticipatory lick count on each trial was calculated as the number of licks during
329 the ISI. Anticipatory lick rates were calculated as the anticipatory lick count divided by the ISI duration.

330 We describe below the trials we analyzed in each study.

- 331 • In [26], we analyzed dopamine activity measured using fluorometry with calcium sensors in ventral
332 striatum (VS). We only analyzed trials to Odor A, the cue that was rewarded identically across all
333 phases, with a reward probability of 75% and an ISI of 3.5 seconds.
- 334 • In [10], dopamine activity was measured using electrophysiology in the ventral tegmental area (VTA).
335 We only analyzed trials to Odor C in the Variable-Expectation task only, which had a reward prob-
336 ability of 90% and an ISI of 2.0 seconds.
- 337 • In [32], dopamine activity was measured using electrophysiology in VTA. We only analyzed trials to
338 Odor A in the high reward probability task, which had a reward probability of 90% and an ISI of 2.0
339 seconds.
- 340 • In [11], dopamine activity was measured using electrophysiology in VTA. We only analyzed trials to
341 Odor B, which had a reward probability of 100% and an ISI of 1.5 seconds.
- 342 • In [33], dopamine activity was measured with fluorometry using calcium sensors (GCaMP) in VTA
343 and dopamine sensors in VS. We analyzed these approaches separately. We only analyzed trials to
344 Odor A, which had a reward probability of 80% and an ISI of 3.0 seconds.

345 **Uncued dopamine peaks**

346 To identify uncued dopamine peaks (Fig 6), we identified time steps during the ITI where the z-scored
347 dopamine activity surpassed 3 standard deviations of its activity. To ensure peaks were defined consistently
348 for each mouse, z-scoring here involved normalizing across all sessions from the same mouse; results were
349 similar when z-scoring per session. To summarize the discriminability of the distributions of lick counts
350 before and after an uncued dopamine peak (Fig 6B,E,F), let the two distributions be indexed by $i = 1$
351 and $i = 2$, with mean μ_i and standard deviation σ_i . Then we calculated the sensitivity index (d') as
352 $d' = (\mu_2 - \mu_1)/\sigma_{1,2}$, where $\sigma_{1,2}^2 = \frac{1}{2}(\sigma_1^2 + \sigma_2^2)$ is the average of the variances.

353 **TD learning simulations**

354 Here we will describe the TD learning procedure used in Fig 5 and Fig 7. Each simulated experiment
355 consisted of concatenated trace conditioning trials for a single cue which was either deterministically
356 rewarded (“CS+”) or unrewarded (“CS-”). The agent’s observations thus consist of the cue, $x_i \in \{0, 1\}$,
357 and the reward, $r_i \in \{0, 1\}$, for all time steps $i = 1, \dots, J$. For simplicity, below we describe x_i and r_i for a
358 single trial t , so that J is the total duration of this trial. The experiment itself is then made up of multiple
359 concatenated trials.

360 Each trial had an intertrial interval (ITI) duration, and an interstimulus interval (ISI) duration. Each
361 ITI was sampled from a Geometric distribution with parameter 0.25, plus a fixed duration of 5 time steps.
362 Each ISI was a constant, $ISI = 8$.

363 In a given trial, $x_i = 1$ when $i = 1 + ITI$, and $x_i = 0$ otherwise. For a CS+ cue, $r_i = 1$ for
 364 $i = 1 + ITI + ISI$, and $r_i = 0$ otherwise. (For a CS- cue, $r_i = 0$ for all i .) Note that $J = 1 + ITI + ISI$.

365 To apply TD learning on trace conditioning trials, where the cue and reward delivery are separated
 366 in time, agents must have a sufficient representation of the cue. Following earlier work [1, 4], we used a
 367 complete serial compound representation, $\mathbf{z}_i \in \mathbb{R}^K$, where $K = 15$, with the first 6 entries corresponding
 368 to time steps within the ITI, the next 8 entries corresponding to time steps within the ISI, and the last
 369 entry serves as a bias term, i.e., $z_i(K) = 1$ for all i . At each time step i , $z_i(k) = 1$ for exactly one k ,
 370 ignoring $k = K$. Thus, \mathbf{z}_i can be thought of as the agent's representation of the current time step within
 371 a given trial. To model sensory noise, we added a noise term $\epsilon_{i,k} \sim \mathcal{N}(0, \sigma_z^2)$ to each $z_i(k)$.

372 TD learning agents used linear value approximation: they learned a weight, $\mathbf{w} \in \mathbb{R}^K$, such that the
 373 agent's value estimate at each time step was $\widehat{V}_i = \mathbf{w}^\top \mathbf{z}_i$. The agent updated \mathbf{w} at each time step using
 374 the reward prediction error (RPE), δ_i . To summarize, after concatenating all trials, TD learning then
 375 proceeded as follows, for all time steps i in the experiment:

$$\begin{aligned} \widehat{V}_i &= \mathbf{z}_i^\top \mathbf{w} \\ \delta_i &= r_i + \gamma \mathbf{z}_{i+1}^\top \mathbf{w} - \mathbf{z}_i^\top \mathbf{w} + b_i + \epsilon_i \\ \mathbf{w} &\leftarrow \alpha_w \mathbf{w} + \alpha \delta_i \mathbf{z}_i \end{aligned} \quad (1)$$

376 where $\gamma = 0.9$ is a discount factor, $b_i = 0$ except during dopamine perturbations (see below), $\epsilon_i \sim \mathcal{N}(0, \sigma^2)$
 377 is i.i.d. Gaussian noise, $\alpha_w = 1$ except when modeling value decay/forgetting, and α is the learning rate.

378 Note that in general we will use i to index time steps, and t to index trials. Also, \widehat{V}_t and δ_t will be
 379 assumed to be the value and RPE, respectively, at the time of CS onset during trial t .

380 Phenotyping

381 To generate model phenotypes (Fig 5A,B,D), we first generated blocks of $N = 10,000$ trace conditioning
 382 trials to a single deterministically rewarded cue. We then trained 500 TD learning agents for both H1 and
 383 H2, where each TD model used different hyperparameters (as defined in Eqn. (1)), sampled uniformly
 384 from the following options: $\alpha \in \{0.001, 0.004, 0.01, 0.02, 0.05, 0.1\}$, $\alpha_w \in \{1 - 10^{-6}, 1 - 10^{-5}, 1 - 10^{-4}\}$,
 385 $\sigma \in \{0.1, 0.2, 0.3, 0.4\}$, and $\alpha_y \in \{0.1, 0.2, \dots, 0.9\}$. Standard agents without sensory noise (Fig 5B) used
 386 $\sigma_z = 0$, while agents with sensory noise (Fig 5D) used $\sigma_z = 0.1$.

387 For each agent, the number of anticipatory licks on a given trial, y_t , was given by:

$$y_t = \alpha_y f(x_t) + (1 - \alpha_y) y_{t-1}$$

388 where $0 < \alpha_y \leq 1$ induces exponential smoothing on y_t , $x_t = \widehat{V}_t$ for agents in H1 (i.e., x_t is CS value),
 389 $x_t = \delta_t$ for agents in H2 (i.e., x_t is the CS RPE), and for simplicity the function f was set to the identity
 390 function, $f(x) = x$.

391 The phenotype of each agent consisted of the Pearson correlation between the number of anticipatory
 392 licks on each trial, y_t , and the CS RPE on trial $t - \tau$, $\delta_{t-\tau}$, for each of $\tau = 0, 1, \dots, 4$.

393 For empirical data (Fig 5C), we used the same phenotyping procedure, where y_t was the actual number
 394 of anticipatory licks on trial t , and δ_t was the CS dopamine activity. Lick counts and CS dopamine were
 395 combined across sessions from all animals after z-scoring per session to control for across-session variability,

396 and padding with NaNs across sessions to ensure that correlations did not compare licking and CS dopamine
397 from different sessions. Significance of correlations was assessed by comparing to the correlations found
398 using trial-shuffled CS DA.

399 **Modeling dopamine perturbations**

400 To model the effects of dopamine manipulations on anticipatory licking in Fig 7, for each trial type we
401 generated 100 random experiments, each consisting of 1,000 trace conditioning trials to a single cue which
402 was either deterministically rewarded (“CS+”) or unrewarded (“CS-”). TD models (Eqn. (1)) were then
403 fit to each of these experiments (separately for each cue) using learning rate $\alpha = 0.1$, and sensory noise
404 $\sigma_z = 0.01$. To model optogenetic excitation of dopamine for [17], we set $b_i = 0.1$ at the time of cue onset.
405 To model optogenetic inhibition of dopamine for [15, 18], we set $b_i = -0.1$ either at the time of cue or
406 reward onset, as indicated by the condition. For all other time steps we set $b_i = 0$. When modeling [17],
407 perturbations were applied during all trials throughout learning, similar to the original experiments. When
408 modeling [15, 18], perturbations were only applied after first training the TD models on 500 baseline trials.

409 Each TD model was then used to generate anticipatory lick rates on each trial t based on either the
410 CS value, V_t , or CS RPE, δ_t , using $p = f(V_t)$ or $p = f(\delta_t)$. For simplicity, and to match what we did in
411 the previous section regarding phenotyping, f was set to the identity function.

412 References

- 413 [1] Richard S Sutton and Andrew G Barto. Time-derivative models of pavlovian reinforcement. In Moore J
414 Gabriel M, editor, *Learning and computational neuroscience: Foundations of adaptive networks*, pages
415 497–537. MIT Press, 1990.
- 416 [2] Elliot A Ludvig, Richard S Sutton, and E James Kehoe. Evaluating the td model of classical condi-
417 tioning. *Learning & behavior*, 40(3):305–319, 2012.
- 418 [3] Samuel J Gershman. A unifying probabilistic view of associative learning. *PLoS Computational*
419 *Biology*, 11(11):e1004567, 2015.
- 420 [4] Wolfram Schultz, Peter Dayan, and P Read Montague. A neural substrate of prediction and reward.
421 *Science*, 275(5306):1593–1599, 1997.
- 422 [5] Jeremiah Y Cohen, Sebastian Haesler, Linh Vong, Bradford B Lowell, and Naoshige Uchida. Neuron-
423 type-specific signals for reward and punishment in the ventral tegmental area. *nature*, 482(7383):85–88,
424 2012.
- 425 [6] Ryunosuke Amo, Sara Matias, Akihiro Yamanaka, Kenji F Tanaka, Naoshige Uchida, and Mitsuko
426 Watabe-Uchida. A gradual temporal shift of dopamine responses mirrors the progression of temporal
427 difference error in machine learning. *Nature neuroscience*, 25(8):1082–1092, 2022.
- 428 [7] Samuel J Gershman, John A Assad, Sandeep Robert Datta, Scott W Linderman, Bernardo L Sabatini,
429 Naoshige Uchida, and Linda Wilbrecht. Explaining dopamine through prediction errors and beyond.
430 *Nature Neuroscience*, 27(9):1645–1655, 2024.
- 431 [8] Hannah M Bayer and Paul W Glimcher. Midbrain dopamine neurons encode a quantitative reward
432 prediction error signal. *Neuron*, 47(1):129–141, 2005.
- 433 [9] Neir Eshel, Michael Bukwich, Vinod Rao, Vivian Hemmelder, Ju Tian, and Naoshige Uchida. Arith-
434 metic and local circuitry underlying dopamine prediction errors. *Nature*, 525(7568):243–246, 2015.
- 435 [10] Neir Eshel, Ju Tian, Michael Bukwich, and Naoshige Uchida. Dopamine neurons share common
436 response function for reward prediction error. *Nature neuroscience*, 19(3):479–486, 2016.
- 437 [11] HyungGoo R Kim, Athar N Malik, John G Mikhael, Pol Bech, Iku Tsutsui-Kimura, Fangmiao Sun,
438 Yajun Zhang, Yulong Li, Mitsuko Watabe-Uchida, Samuel J Gershman, et al. A unified framework
439 for dopamine signals across timescales. *Cell*, 183(6):1600–1616, 2020.
- 440 [12] Elizabeth E Steinberg, Ronald Keiflin, Josiah R Boivin, Ilana B Witten, Karl Deisseroth, and Pa-
441 tricia H Janak. A causal link between prediction errors, dopamine neurons and learning. *Nature*
442 *Neuroscience*, 16(7):966–973, 2013.
- 443 [13] Chun Yun Chang, Guillem R Esber, Yasmin Marrero-Garcia, Hau-Jie Yau, Antonello Bonci, and
444 Geoffrey Schoenbaum. Brief optogenetic inhibition of dopamine neurons mimics endogenous negative
445 reward prediction errors. *Nature Neuroscience*, 19(1):111–116, 2016.

- 446 [14] Benjamin T Saunders, Jocelyn M Richard, Elyssa B Margolis, and Patricia H Janak. Dopamine
447 neurons create Pavlovian conditioned stimuli with circuit-defined motivational properties. *Nature*
448 *neuroscience*, 21(8):1072–1083, 2018.
- 449 [15] Kwang Lee, Leslie D Claar, Ayaka Hachisuka, Konstantin I Bakhurin, Jacquelyn Nguyen, Jeremy M
450 Trott, Jay L Gill, and Sotiris C Masmanidis. Temporally restricted dopaminergic control of reward-
451 conditioned movements. *Nature neuroscience*, 23(2):209–216, 2020.
- 452 [16] Johann Du Hoffmann and Saleem M Nicola. Dopamine invigorates reward seeking by promoting
453 cue-evoked excitation in the nucleus accumbens. *Journal of Neuroscience*, 34(43):14349–14364, 2014.
- 454 [17] Joachim Morrens, Çağatay Aydin, Aliza Janse van Rensburg, José Esquivelzeta Rabell, and Sebastian
455 Haesler. Cue-evoked dopamine promotes conditioned responding during learning. *Neuron*, 106(1):142–
456 153, 2020.
- 457 [18] Ruud Van Zessen, Jacques P Flores-Dourojeanni, Timon Eekel, Siem van den Reijen, Bart Lodder,
458 Azar Omrani, Marten P Smidt, Geert MJ Ramakers, Geoffrey van der Plasse, Garret D Stuber, et al.
459 Cue and reward evoked dopamine activity is necessary for maintaining learned pavlovian associations.
460 *Journal of Neuroscience*, 41(23):5004–5014, 2021.
- 461 [19] David M Egelman, Christophe Person, and P Read Montague. A computational role for dopamine
462 delivery in human decision-making. *Journal of Cognitive Neuroscience*, 10(5):623–630, 1998.
- 463 [20] Samuel M McClure, Nathaniel D Daw, and P Read Montague. A computational substrate for incentive
464 salience. *Trends in neurosciences*, 26(8):423–428, 2003.
- 465 [21] Michael J Frank. Dynamic dopamine modulation in the basal ganglia: a neurocomputational account
466 of cognitive deficits in medicated and nonmedicated parkinsonism. *Journal of cognitive neuroscience*,
467 17(1):51–72, 2005.
- 468 [22] Peter Dayan and Bernard W Balleine. Reward, motivation, and reinforcement learning. *Neuron*,
469 36(2):285–298, 2002.
- 470 [23] Rafal Bogacz. Dopamine role in learning and action inference. *Elife*, 9:e53262, 2020.
- 471 [24] Charles R Gerfen and D James Surmeier. Modulation of striatal projection systems by dopamine.
472 *Annual review of neuroscience*, 34(1):441–466, 2011.
- 473 [25] Asha K Lahiri and Mark D Bevan. Dopaminergic transmission rapidly and persistently enhances
474 excitability of d1 receptor-expressing striatal projection neurons. *Neuron*, 106(2):277–290, 2020.
- 475 [26] Lechen Qian, Mark Burrell, Jay A Hennig, Sara Matias, Venkatesh N Murthy, Samuel J Gershman, and
476 Naoshige Uchida. Prospective contingency explains behavior and dopamine signals during associative
477 learning. *Nature neuroscience*, pages 1–13, 2025.
- 478 [27] Andrew T Marshall, Briac Halbout, Angela T Liu, and Sean B Ostlund. Contributions of pavlovian
479 incentive motivation to cue-potentiated feeding. *Scientific reports*, 8(1):2766, 2018.

- 480 [28] SD Paolo. Dopamine on d2-like receptors “reboosts” dopamine d1-like receptor-mediated behavioural
481 activation in rats licking for sucrose. *Neuropharmacology*, 58(7):1085–1096, 2010.
- 482 [29] SD Paolo. Licking microstructure in response to novel rewards, reward devaluation and dopamine
483 antagonists: possible role of d1 and d2 medium spiny neurons in the nucleus accumbens. *Neuroscience*
484 *& Biobehavioral Reviews*, page 105861, 2024.
- 485 [30] Koji Toda, Nicholas A Lusk, Glenn DR Watson, Namsoo Kim, Dongye Lu, Haofang E Li, Warren H
486 Meck, and Henry H Yin. Nigrotectal stimulation stops interval timing in mice. *Current Biology*,
487 27(24):3763–3770, 2017.
- 488 [31] Alam Coss, Ernesto Suaste, and Ranier Gutierrez. Lateral nac shell d1 and d2 neuronal ensembles
489 concurrently predict licking behavior and categorize sucrose concentrations in a context-dependent
490 manner. *Neuroscience*, 493:81–98, 2022.
- 491 [32] Hideyuki Matsumoto, Ju Tian, Naoshige Uchida, and Mitsuko Watabe-Uchida. Midbrain dopamine
492 neurons signal aversion in a reward-context-dependent manner. *Elife*, 5:e17328, 2016.
- 493 [33] Ryunosuke Amo, Naoshige Uchida, and Mitsuko Watabe-Uchida. Glutamate inputs send prediction
494 error of reward, but not negative value of aversive stimuli, to dopamine neurons. *Neuron*, 112(6):1001–
495 1019, 2024.
- 496 [34] Martin Darvas, Amanda M Wunsch, Jeffrey T Gibbs, and Richard D Palmiter. Dopamine depen-
497 dency for acquisition and performance of pavlovian conditioned response. *Proceedings of the National*
498 *Academy of Sciences*, 111(7):2764–2769, 2014.
- 499 [35] Jun Zhang, Kent C Berridge, Amy J Tindell, Kyle S Smith, and J Wayne Aldridge. A neural compu-
500 tational model of incentive salience. *PLoS Computational Biology*, 5(7):e1000437, 2009.
- 501 [36] Kent C Berridge and Terry E Robinson. What is the role of dopamine in reward: hedonic impact,
502 reward learning, or incentive salience? *Brain Research Reviews*, 28(3):309–369, 1998.
- 503 [37] Benjamin T Saunders and Terry E Robinson. The role of dopamine in the accumbens core in the
504 expression of pavlovian-conditioned responses. *European Journal of Neuroscience*, 36(4):2521–2532,
505 2012.
- 506 [38] Amanda G Iglesias, Alvin S Chiu, Jason Wong, Paolo Campus, Fei Li, Jasmine K Bhatti, Shiv A
507 Patel, Karl Deisseroth, Huda Akil, Christian R Burgess, et al. Inhibition of dopamine neurons pre-
508 vents incentive value encoding of a reward cue: With revelations from deep phenotyping. *Journal of*
509 *Neuroscience*, 43(44):7376–7392, 2023.
- 510 [39] Natalie L Johnson, Anamaria Cotelso-Larrea, Lucas A Stetzik, Umit M Akkaya, Zihao Zhang, Marie A
511 Gadziola, Adrienn G Varga, Minghong Ma, and Daniel W Wesson. Dopaminergic signaling to ventral
512 striatum neurons initiates sniffing behavior. *Nature communications*, 16(1):336, 2025.
- 513 [40] Charltien Long, Kwang Lee, Long Yang, Theresia Dafalias, Alexander K Wu, and Sotiris C Mas-
514 manidis. Constraints on the subsecond modulation of striatal dynamics by physiological dopamine
515 signaling. *Nature neuroscience*, 27(10):1977–1986, 2024.

- 516 [41] Zhaorong Chen, Zhi-Yu Zhang, Wen Zhang, Taorong Xie, Yaping Li, Xiao-Hong Xu, and Haishan Yao.
517 Direct and indirect pathway neurons in ventrolateral striatum differentially regulate licking movement
518 and nigral responses. *Cell reports*, 37(3), 2021.
- 519 [42] Irene A Yun, Saleem M Nicola, and Howard L Fields. Contrasting effects of dopamine and glutamate
520 receptor antagonist injection in the nucleus accumbens suggest a neural mechanism underlying cue-
521 evoked goal-directed behavior. *European Journal of Neuroscience*, 20(1):249–263, 2004.
- 522 [43] Briac Halbout, Andrew T Marshall, Ali Azimi, Mimi Liljeholm, Stephen V Mahler, Kate M Wassum,
523 and Sean B Ostlund. Mesolimbic dopamine projections mediate cue-motivated reward seeking but
524 not reward retrieval in rats. *Elife*, 8:e43551, 2019.
- 525 [44] Sarah Fischbach-Weiss, Rebecca M Reese, and Patricia H Janak. Inhibiting mesolimbic dopamine
526 neurons reduces the initiation and maintenance of instrumental responding. *Neuroscience*, 372:306–
527 315, 2018.
- 528 [45] Abigail Kalmbach, Vanessa Winiger, Nuri Jeong, Arun Asok, Charles R Gallistel, Peter D Balsam,
529 and Eleanor H Simpson. Dopamine encodes real-time reward availability and transitions between
530 reward availability states on different timescales. *Nature communications*, 13(1):3805, 2022.
- 531 [46] Anja Lex and Wolfgang Hauber. Dopamine d1 and d2 receptors in the nucleus accumbens core and
532 shell mediate pavlovian-instrumental transfer. *Learning & memory*, 15(7):483–491, 2008.
- 533 [47] Joaquim Alves Da Silva, Fatuel Tecuapetla, Vitor Paixão, and Rui M Costa. Dopamine neuron activity
534 before action initiation gates and invigorates future movements. *Nature*, 554(7691):244–248, 2018.
- 535 [48] Allison E Hamilos, Giulia Spedicato, Ye Hong, Fangmiao Sun, Yulong Li, and John A Assad. Slowly
536 evolving dopaminergic activity modulates the moment-to-moment probability of reward-related self-
537 timed movements. *Elife*, 10:e62583, 2021.
- 538 [49] Allison E Hamilos, Isabella C Wijsman, Qinxin Ding, Pichamon Assawaphadungsit, Zeynep Ozcan,
539 Elias Norri, Kimberly Reinhold, Bernardo L. Sabatini, and John A Assad. Dopamine reward transients
540 calibrate movement timing via one-shot updates to behavioral vigor. *bioRxiv*, 2025.
- 541 [50] Laurens Winkelmeier, Carla Filosa, Renée Hartig, Max Scheller, Markus Sack, Jonathan R Reinwald,
542 Robert Becker, David Wolf, Martin Fungisai Gerchen, Alexander Sartorius, et al. Striatal hub of
543 dynamic and stabilized prediction coding in forebrain networks for olfactory reinforcement learning.
544 *Nature Communications*, 13(1):3305, 2022.
- 545 [51] CR Gallistel and John Gibbon. Time, rate, and conditioning. *Psychological Review*, 107(2):289–344,
546 2000.
- 547 [52] Charles Randy Gallistel. Reconceptualized associative learning. *Perspectives on Behavior Science*,
548 48(2):203–239, 2025.
- 549 [53] Samuel J Gershman. Bridging computation and representation in associative learning. *Computational*
550 *Brain & Behavior*, pages 1–15, 2025.

- 551 [54] Justin A Harris and Charles Randy Gallistel. Information, certainty, and learning. *eLife*, 13:RP102155,
552 2026.
- 553 [55] Robert A Rescorla. Probability of shock in the presence and absence of CS in fear conditioning.
554 *Journal of Comparative and Physiological Psychology*, 66(1):1, 1968.